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54 Method of detecting AIDS virus infection.

57 A method is disclosed for detecting the presence of HTLV-III infected cells in a medium. The method comprises contacting the medium with monoclonal antibodies against an antigen produced as a result of the infection and detecting the binding of the antibodies to the antigen. The antigen may be a gene product of the HTLV-III virus or may be bound to such gene product. On the other hand the antigen may not be a viral gene product but may be produced as a result of the infection and may further be bound to a lymphocyte. The medium may be a human body fluid or a culture medium. A particular embodiment of the present method involves a method for determining the presence of an AIDS virus in a person. The method comprises combining a sample of a body fluid from the person with a monoclonal antibody that binds to an antigen produced as a result of the infection and detecting the binding of the monoclonal antibody to the antigen. The presence of the binding indicates the presence of an AIDS virus infection. Also disclosed are novel monoclonal antibodies, novel compositions of matter, and novel diagnostic kits.

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METHOD OF DETECTING AIDS VIRUS INFECTION

The disease called Acquired Immune Deficiency Syndrome (AIDS) has been recognized only recently but has created an alarming situation in many parts of the world. The number of diagnosed cases per year has escalated rapidly and a large number of deaths has resulted. Until recently, the disease apparently has been prevalent primarily in homosexual men with multiple sexual partners, intravenous drug users, hemophiliacs, blood transfusion recipients, and close heterosexual contacts of members of the above group. Despite the mounting concern with regard to AIDS, the origin and cure of the disease have alluded the medical and scientific community although a great deal of research has already been carried out with respect to the disease.

A major advance and understanding of the basis of AIDS has been the isolation of a novel class of retrovirus from patients suffering from AIDS or AIDS-related complex (ARC). This group of viruses is variously called lymphadenopathy virus (LAV) as referred to by S. Wain Hobson, et al., *Cell*, 40, 9-17 (1978), human T lymphotropic virus III (HTLV-III) as described by L. Ratner, et al., *Nature*, 313, 277-84 (1985) and AIDS-associated retrovirus (ARV) as disclosed by R. Sanchez Pescador, et al., *Science*, 227, 484-492 (1985). From a morphologic and biologic standpoint, this group of viruses is most closely related to the lentivirus family.

The primary targets of affliction of the AIDS virus in the human body are specific subpopulations of T-cells. The severe immune deficiency of these patients results from an unusually low proportion of T-cells (T4) in their lymphocyte population. As a result the availability of many T4 helper functions is reduced. One of such functions is the production of antibodies by the B-cells.

The pronounced depression of cellular immunity that occurs in patients with AIDS and the quantitative modifications of subpopulations of their T-lymphocytes suggests that T-cells or a subset of T-cells might be a preferential target for the putative infectious agent. Alternatively, these modifications may result from subsequent infections. The depressed cellular immunity may result in serious opportunistic infections in AIDS patients, many of whom develop Kaposi's Sarcoma. Multiple lymphadenopathies have also been described in homosexual males and infants who may or may not develop AIDS. The lymphadenopathies may correspond to an early or milder form of the disease.

There are currently three major tests for establishing the presence of serum antibody specific for HTLV-III virus. The most widely employed procedure is the ELISA screen which uses either wells or beads coated with HTLV-III viral lysate. Specific antibody from a seropositive specimen binds to the antigen coated surface, followed by an enzyme linked, anti-human antibody and appropriate chromogenic substrate.

The second antibody detection procedure, most often used as a confirmatory test for the ELISA screen, is the enzyme-linked immunoelectrotransfer blot or Western Blot technique. An HTLV-III viral lysate is resolved by SDS-Polyacrylamide Gel Electrophoresis (SDS-PAGE) into a series of bands, each defining a particular product of the viral genome. The antigen bands are then transblotted to a nitrocellulose membrane for use in an enzyme-linked immunosorbent assay procedure, similar to the ELISA screen. The principle advantage of the Western Blot is the resolution of the viral antigens into discreet bands providing greatly enhanced specificity. There are two bands elucidated by the Western Blot, either or both of which are considered diagnostic for HTLV-III specific antibody when detected by the ELISA portion of the test. The first band is the 24,000 dalton core protein. The second band is a 41,000 dalton glycoprotein which makes up a portion of the viral envelope. Equivocal results in the ELISA screen are frequently resolved by the Western Blot. Also, positive screens (especially among low risk group individuals) will often prove to be negative when reevaluated by the Western Blot.

The third antibody test is an indirect immunofluorescence assay using virus infected lymphoid cells which are reacted first with patient serum and then a fluorochrome-conjugated anti-human antibody. Fluorescence of the infected cells constitutes a positive test. The use of uninfected lymphoid cells provides a negative control which would detect antibody specific for the cells and not the virus.

The use of the ELISA screen and Western Blot confirmation tests for HTLV-III antibody has been a valuable screening tool for the elimination of contaminated blood products from the national blood supply as well as detection of infectious organ and sperm donors. However, the detection of serum antibody specific for HTLV-III virus is not a diagnosis of AIDS. Seropositivity does not prove that the patient has been exposed to live virus, is infectious, or has actively growing virus, nor does it provide any indication concerning the course of the disease. A significant percentage of seropositive hemophiliacs have failed to

show the presence of virus in their blood by current techniques. Presumably, these individuals have been exposed to inactivated virus contained in the heat-treated factor VIII preparations. However, in other seropositive individuals the correlation between the presence of specific antibody and the ability to culture HTLV-III virus is strong.

5 The need for practical procedures for detecting the presence of antigens associated with HTLV-III viral infection in human body fluids is essential for a reliable diagnostic test for AIDS as well as for providing the means to evaluate the effect of several experimental drugs now being tested for their efficacy in treating AIDS victims.

10 Currently, the principle method for detecting HTLV-III virus in blood involves mixing patient lymphocytes with phytohemagglutinin (PHA)-stimulated peripheral blood lymphocytes (PBLs) with the addition of fresh PHA-stimulated PBLs every three days. Supernatant samples are then tested every three days for the presence of reverse transcriptase. Cells are evaluated by electron microscopy for the presence of virus. Clearly, this method is restricted to facilities with high level, P3 containment as well as sophisticated tissue culture and analysis capabilities. Results may take several weeks and the reliability of infecting lymphocyte
15 cultures with virus is questionable. Use of a cell line known as H9 has helped some but most of the above problems still exist.

Certain diagnostic techniques for determining the presence of AIDS or AIDS antibodies are known. One such technique is a helper-suppressor cell assay performed on T-cells and T-cell subpopulations. The reversal of the helper-suppressor ratio is taken as an indication of the possible presence of AIDS virus. This
20 assay has low specificity and sensitivity. Another technique involves a determination of the thymosin level, the elevation of the level being an indication of the presence of AIDS. This technique gives positive results only when the disease has reached an advanced stage. Furthermore, it has been reported that the technique fails to give consistently positive readings in all cases when AIDS is known to be present. The diagnosis of AIDS in mammalian subjects involving the use of immunocyto adherents (rosette inhibition)
25 techniques is disclosed in European Patent Application Publication No. 0154499. U.S. Patent No. 4,520,113 discloses the serological detection of antibodies to HTLV-III in sera of patients with AIDS and pre-AIDS conditions.

A late differentiation antigen associated with the helper inducer function of human T-cells is described in Nature (1985) 318:465-467. The diagnosis of AIDS or lymphadenopathic syndrome using specified T-
30 lymphotropic retrovirus antigen is discussed in European Patent Application Publication No. 0138667.

The present invention is directed to a method for detecting the presence of HTLV-III infected cells in a medium such as a body fluid or a culture medium. The method comprises contacting the medium with a monoclonal antibody against an antigen produced as a result of the infection and detecting the binding of
35 the antibodies of the the antigen. The antigen may be HTLV-III viral gene product or it may be bound to such a product. Alternatively, the antigen may be produced as a result of the infection and found on a lymphocyte such as a T-helper cell.

The invention further includes an immunoassay method for the detection of an AIDS virus related infection in a host. The method comprises combining a body fluid from the host with a monoclonal antibody that binds to an antigen that is present only when the AIDS virus infection is present and examining the
40 combination for the presence of immune complexes comprising the antigen and the monoclonal antibody.

The invention also includes an antibody capable of binding to a serum antigen produced as a result of a human HTLV-III viral infection. The antigen may be on an HTLV-III viral gene product or it may be bound to an HTLV-III viral gene product. Alternatively, the antigen may be bound to a lymphocyte as a result of an HTLV-III virus infection. Hybrid continuous cell lines having the identifying characteristic of expressing the
45 antibodies of the invention are also included.

Diagnostic kits are also provided and comprise in a packaged combination a monoclonal antibody capable of binding to an antigen produced as a result of an infection by the viral causative agent of AIDS or a labeled derivative of such antibody.

Another aspect of the invention concerns a method for the early detection of HTLV-III viral infection in
50 cell cultures. The method comprises combining cells suspected of HTLV-III viral infection with a monoclonal antibody of the invention. Detection of binding of the antibody and the antigen indicates the presence of HTLV-III viral infection in the cell culture.

The invention also includes compositions of matter comprising a complex of a monoclonal antibody that binds to (a) an HTLV-III gene produce or a derivative thereof and such (b) gene product or derivative.
55 Another composition of matter in accordance with the present invention comprises a complex of (a) a monoclonal antibody capable of binding to a cell bound antigen other than an HTLV-III gene product and produced as a result of a HTLV-III infection and (b) such antigen.

Before proceeding further with a description of the specific embodiments of the present invention, a number of terms will be defined.

"AIDS virus" means HTLV-III, ARV, LAV and related viruses known to cause or suspected to cause AIDS, ARC, LAD, and related disorders.

5 "AIDS related diseases" means a disease which is AIDS, AIDS related complex (ARC), lymphadenopathy (LAD), progressive generalized lymphadenopathy (PGL), and related disorders that are known or suspected to be caused by an AIDS virus infection.

"Soluble AIDS antigen" means an antigen produced as a result of an AIDS virus infection not bound to a cell membrane and found characteristically in people that have been infected with an AIDS related virus.

10 "Cell-bound AIDS antigen" means an antigen bound to a cell membrane and found characteristically in people that have been infected with an AIDS related virus.

"Body fluid" means a liquid or semi-solid material obtained from the body of a mammalian subject. The liquid material may be sterile or nonsterile and usually contains cells. The liquid material may be employed without further treatment or the liquid material may be treated to remove cells, debris, and the like.
15 Exemplary of body fluids are whole blood, lymphatic fluid, serum, plasma, saliva, semen, and cerebral spinal fluid. Body fluid may be removed from the subject, for example, by means of a syringe or needle or by natural expulsion.

"Cell" means a unicellular organism such as a bacterium and also refers to normal and transformed cells, usually mammalian, obtained from a body fluid. Cells may be infected or noninfected, or may be
20 cultured or not cultured.

"Lymphocyte" means a variety of white blood corpuscles or cells which arise in the reticular tissue of the lymph glands and lymph nodes. The nucleus is single and is surrounded by protoplasm which is generally described as nongranular. There are small lymphocytes, which are about the size of a red corpuscle and large lymphocytes which are probably lymphocytes in their developing stage and are two or
25 three times larger than the small lymphocytes and contain a larger proportion of protoplasm.

"T lymphocyte" means those lymphocytes originating in the thymus. The main function of T cells is regulation of the immune response of B cells via helper and suppressor functions. T cells do not make conventional antibodies and thus differ from antibody forming cells arising from the bone marrow which are designated as B lymphocytes.

30 "T-helper cell" means a lymphocyte which has a main function of regulating the immune response of B cells in a help capacity as opposed to a suppressor capacity.

"Viral gene product" means a product such as an antigen derived from the AIDS related virus by an RNA or DNA replication.

The present invention is directed to a method for detecting the presence of cells infected with an AIDS
35 related virus such as HTLV-III in a medium. The method comprises contacting the medium with monoclonal antibodies against an antigen produced as a result of the infection and further detecting the binding of the antibodies to the antigen. If binding between the antigen and antibodies occurs, the presence of an AIDS related virus infection in the cells in the medium is indicated. The medium can be for example a body fluid or a culture medium. The antigen may be a soluble antigen or a cell bound antigen. Furthermore, the
40 antigen may be a viral gene product, or bound to a viral gene product. Where the antigen is not a viral gene product it is produced as a result of the infection of cells with the AIDS related virus. The antigen may be bound to a lymphocyte such as for example a T-helper cell or to a macrophage. Usually, where the antigen is a viral gene product, the method involves detection of the binding of antibodies to the infected cells.

An antibody included within the scope of the present invention is capable of binding to a serum antigen
45 produced as a result of an HTLV-III viral infection. The antigen is an HTLV-III viral gene product or is bound to a HTLV-III viral gene product. Alternatively, the antigen may be bound to a lymphocyte and is produced as a result of the HTLV-III infection. Usually, where the antigen is soluble it is not a viral gene product.

Monoclonal antibodies useful in the method of the invention may be produced according to the standard techniques of Köhler and Milstein, *Nature* 265:495-497, 1975. For example, partially purified viral
50 lysates derived from the growth of HTLV-III in the human lymphoma, H9 cell line can be used as the immunogen. Samples of the HTLV-III antigen preparations are injected into a mouse and, after a sufficient time, the mouse is sacrificed and spleen cells obtained. The spleen cell chromosomes encoding the base sequences for the desired immunoglobulins are immortalized by fusing the spleen cells, to myeloma cell lines, generally in the presence of a non-ionic detergent, for example, polyethylene glycol. The resulting
55 cells, which include the fused hybridomas, are allowed to grow in a selective medium, such as HAT-medium, and the surviving cells are grown in such medium using limiting dilution conditions. The cells are grown in a suitable container, .g., microtiter wells, and the supernatant is screened for monoclonal antibodies having the desired specificity.

Various techniques exist for enhancing yields of monoclonal antibodies, such as injection of the hybridoma cells into the peritoneal cavity of a mammalian host, which accepts the cells, and harvesting the ascites fluid. Where an insufficient amount of the monoclonal antibody collects in the ascites fluid, the antibody is harvested from the blood of the host. Various conventional ways exist for isolation and purification of the monoclonal antibodies from other proteins and other contaminants (see KÖhler and Milstein, supra).

One such monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 3D8. This monoclonal antibody binds to determinants on antigens that are HTLV-III gene products bound to a cell. Lysates of AIDS infected cells are submitted to sodium dodecylsulphate-polyacrylamide one-dimensional gel electrophoresis (SDS-PAGE). Monoclonal antibody bound to nitrocellulose was developed with the SDS-PAGE gel according to Western Blot techniques. It was determined that the antibody 3D8 binds to a reverse transcriptase of 55 kilodaltons and 65 kilodaltons (p55 and p65), a core protein of 24 kilodaltons (p24), a glycoprotein envelope antigen of 41 kilodaltons (gp41), and one undefined protein of 39 kilodaltons, (p39). The antigens designated p55 and p65, the core protein antigen p24, and the glycoprotein envelope antigen gp41 have been described in Science (1986) 231:1289-1291. Undefined proteins p28, p39, p49 and p52 may be antigens which are bound to a cell but are not HTLV-III gene products. These antigens may be produced as a result of an AIDS viral infection and identify a cell as infected. On the other hand the p28, p49 and p52 proteins may be viral gene products that are not immunogenic in man. The p41, p55, and p65 antigens appear to be soluble antigens. Based on Western Blot observations the 3D8 antibody and the antibodies described below may recognize different determinants on the above described antigens. The 3D8 antibody is of the IgM isotype. The 3D8 antibody does not bind to normal lymphocytes, or cells from cell lines H9 or CEM.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 3F7. A similar determination employing SDS-PAGE and Western Blot techniques indicated that antibody 3F7 binds to the p24 antigen, the p49 antigen, the p55 antigen, and the p65 antigen. The 3F7 antibody is of the IgG 1 isotype. The 3F7 antibody does not bind with high intensity to normal lymphocytes, H9 cells or CEM cells. By the term "high intensity" is meant that peaks for stained cells and non-stained cells in flow cytometry are separated.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 3G12. By a combination SDS-PAGE and Western Blot techniques it was determined that antibody 3G12 binds to the p24 antigen and the p55 antigen. The 3G12 antibody is of the IgG 1 isotype. The 3G12 antibody does not bind with high intensity to normal lymphocytes or CEM cells.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 2A11. Antibody 2A11 binds to the p24 antigen, the p39 antigen, and the p65 antigen. The 2A11 antibody is an IgM. The 2A11 antibody does not bind with high intensity to normal lymphocytes, H9 or CEM cells.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 4G2. As determined by SDS-PAGE and Western Blot techniques the 4G2 antibody binds to the p24 antigen and the p55 antigen. The 4G2 antibody is of the IgM isotype. The 4G2 antibody does not bind with high intensity to normal lymphocytes, H9 or CEM cells.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 1E2. This monoclonal antibody binds to the p55 antigen and the p49 antigen. The 1E2 antibody is of the IgM isotype. The 1E2 antibody does not bind with high intensity to normal lymphocytes or CEM cells.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 3G8. Antibody 3G8 binds to the p24 antigen, the p39 antigen and the p41 antigen. The 3G8 antibody is an IgM isotype. The 3G8 antibody does not bind with high intensity to normal lymphocytes, H9 cells or CEM cells.

Another monoclonal antibody useful in the method of the present invention is exemplified by a novel antibody expressed by the hybrid continuous cell line designated 1C11. Antibody 1C11 binds to the p55 antigen and p65 antigen. The 1C11 antibody is an IgM. Cell line 1C11 was obtained by re-cloning cell line 3D8. Apparently, cell line 3D8 is a mixture of two cell lines, namely, 3G8 and 1C11. When individual cells from 3D8 are isolated and grown, two cell lines, 3G8 and 1C11, are obtained. The combined Western Blot patterns for 3G8 and 1C11 are superimposable on the Western Blot pattern of 3D8.

Also included within the scope of the invention and the definition of monoclonal antibody are useful binding fragments of the monoclonal antibody such as Fab, F(ab')₂, Fv, and so forth. The antibody fragments are obtained by conventional techniques. For example, useful binding fragments may be prepared by peptidase digestion of the antibody using papain or pepsin.

5 While the above specific examples of the novel antibodies of the invention are directed to antibodies of the IgG and IgM classes from a murine source, this is not meant to be a limitation. The above antibodies and those antibodies having functional equivalency with the above antibody, whether from a murine source, other mammalian source including human, rat, or other sources, or combinations thereof are included within the scope of this invention, as well as other classes such as IgA, IgE, and the like, including isotypes within
10 such classes and also other isotypes within the IgG and IgM classes. By the term "functional equivalency" is meant that the antibody is capable of binding to the above-described antigen and capable of competing with an antibody of the invention for binding to such antigen. That is, such antibody, when combined with a sample containing such antigen, will bind to such antigen and will block an antibody of the invention from binding to such antigen. Mammalian (e.g. human) monoclonal antibodies can be prepared by techniques
15 described in Olsson et al. Proc. Nat. Acad. Sci. U.S.A. (1980)77:5429-5431; Engelman et al., "Human Hybridomas and Monoclonal Antibodies", Plenum Press, N.Y., N.Y. (1985). The fusion techniques are similar to those described above for the preparation of murine hybridomas. The following cell lines are exemplary of those which can be utilized in preparing the hybrid continuous cell lines: NS-1 (American Type Culture Collection, Rockville, Maryland, ATTC), SP-2 (ATTC), HL-1 (Ventrex, Portland, Maine).

20 In carrying out a method in accordance with the present invention, a sample of a body fluid is secured from the subject and prepared as follows: For preparation of lymphocytes mononuclear cells are separated from the rest of the blood. For example peripheral blood is anticoagulated with heparin, diluted with an equal volume of phosphate buffer and subjected to density gradient centrifugation, thereby separating the mononuclear cells (primarily lymphocytes) from the rest of the blood. The cells are then washed in buffer
25 and suspended in buffer with a small amount, (e.g., 0.01-1%, preferably about 0.1%), gelatin and a small amount, (e.g., 0.01-1%, preferably about 0.1%), sodium azide at 5×10^6 cells per ml. Alternatively, separation of the white blood cells can be achieved by whole blood hemolysis.

For preparation of serum, peripheral blood is drawn into a plain glass tube, allowed to clot and then centrifuged to separate the serum from the clot.

30 The amount of sample will depend on the nature of the particular assay method employed and the particular body fluid to be analyzed. The following sample amounts are provided by way of example and not limitation; those skilled in the art will be able to broadly practice the method of the present invention on the basis of the description provided herein. For serum or plasma samples the amount of sample generally will be in the range of about 0.1 ml to 5.0 ml, preferably 1.0 ml to 2.0 ml. For semen samples, the amount
35 will generally be about 0.05 ml to 1.0 ml, preferably 0.1 to 0.5 ml.

The sample is next contacted with one of the aforementioned antibodies or a combination thereof under conditions for binding of the antibody to the antigen in the sample to form immune, i.e., antigen-antibody complexes. Depending on the nature of the assay, the antibody can be bound to a support or can be in an aqueous medium. The contact between the antibody and the sample is generally carried out in an aqueous
40 buffered medium. The buffers which can be employed include phosphate, tris, bicarbonate, etc. The pH employed depends on the nature of the sample and the antibody, and is generally in the range of from about 5 to 8. The aqueous medium may additionally contain organic polar solvents in an amount of from about 0 to 40%. The organic polar solvents are water soluble and generally have from about 1 to 10 carbon atoms and from about 1 to 4 oxygen atoms. The antibody concentration will also depend on the nature of
45 the assay and the volume of the sample to be tested. The amount of antibody will generally be in the 0.5 μ g to 2 μ g per ml range.

After the period of contact between the sample and the antibody and depending on the nature of the assay, the combination may be treated to remove unreacted antibody. This can be accomplished by washing with an aqueous, usually buffered, medium. In general, the amount of wash solution should be
50 sufficient to remove the unreacted antibody. The number and types of washing steps depend on whether a heterogeneous or homogeneous assay approach is employed.

Next, the presence of binding of the antibody to the antigen in the sample, which binding is an indication of the presence of an AIDS-related virus infection in the subject, is observed. That is, the number of antigen-antibody (immune) complexes formed is determined. To make the determination of the presence
55 of binding, means for producing a detectable signal in relation to the presence of antigen in the sample is incorporated into the assay system. For example, one may conjugate the antibody employed in the assay to a label which is a substance capable of producing a detectable signal in relation to the presence or absence of an AIDS-related viral infection. The label can be, for example, an enzyme, radioisotope, particle,

support, chromogen, chemiluminescer, fluoescer, coenzyme, free radical, or bacteriophage. The number of labels employed for the antibody is generally determined by the requirements of the method of the present invention and the availability of sites for linking the label to the antibody. Methods for linking labels to antibodies are known in the art. See for example, U.S. Patent Nos. 4,720,450; 4,235,869; 3,935,074; and 3,996,345.

Alternatively, one may employ a labeled specific binding partner for the antibody, which may be, for example, a labeled antibody specific for the antibody employed or labeled antigen that binds to the antibody. Where the monoclonal antibody is derived from a murine source, a labeled anti-mouse immunoglobulin specific for the antibody employed in the method may be used. Such immunoglobulins may be raised according to standard techniques by injecting a suitable host with the monoclonal antibody, waiting for an appropriate time, and harvesting the anti-mouse immunoglobulins from the blood of the injected host.

The method and antibodies of the invention may be adapted to most assays involving antigen-antibody reactions. The assays may be homogeneous or heterogeneous. In a homogeneous assay approach, the sample may be pretreated if necessary to remove unwanted materials. The immunological reaction usually involves the specific antibody, a labeled analyte, and the sample of interest. The signal arising from the label is modified, directly or indirectly, upon the binding of the antibody to the labeled analyte. Both the immunological reaction and detection of the extent thereof are carried out in a homogeneous solution. Immunochemical labels which may be employed include free radicals, fluorescent dyes, enzymes, bacteriophages, coenzymes, and so forth. Exemplary of homogeneous assays are the enzyme modulated immunoassay technique (EMIT®) described in U.S. Patent No. 3,817,837.

In a heterogeneous assay approach, the reagents are usually the sample, the antibody, and means for producing a detectable signal. The sample is generally placed on a support, such as a plate or a slide, and contacted with the antibody in an aqueous phase. The support is then separated from the liquid phase and either the support phase or the liquid phase is examined for a detectable signal employing means for producing such signal. Means for producing a detectable signal in heterogeneous assays includes the use of radioactive labels, fluoescers, enzymes, and so forth. Exemplary of heterogeneous immunoassays are the radioimmunoassay (RIA), immunofluorescence methods, enzyme-linked immunoassays, such as the enzyme-linked immunosorbent assay (ELISA, see U.S. Patent Nos. 3,654,090; 3,839,153; 3,850,752; 4,016,043; and Reissue Patent No. 29,169.

For a more detailed discussion of the above immunoassay techniques, see "Enzyme-Immunoassay," by Edward T. Maggio, CRC Press, Inc., Boca Raton, Florida, 1980. See also, for example, U.S. Patent Nos. 3,690,834; 3,791,932; 3,817,837; 3,850,578; 3,853,987; 3,867,517; 3,901,654; 3,935,074; 3,984,533; 3,996,345; and 4,098,876, which listing is not intended to be exhaustive.

A particular embodiment of an assay in accordance with the present invention, by way of illustration and not limitation, involves the use of a support such as a slide or a well of a petri dish. The technique involves fixing the sample to be analyzed on the support with an appropriate fixing material such as acetone and incubating the sample on the slide with a monoclonal antibody of the invention. After washing with an appropriate buffer such as, for example, phosphate buffered saline, the support is contacted with a labeled specific binding partner for the monoclonal antibody. After incubation as desired, the slide is washed a second time with an aqueous buffer and the determination is made of the binding of the monoclonal antibody to the sample. If the label is fluorescent, the slide may be covered with a fluorescent antibody mounting fluid on a cover slip and then examined with a fluorescent microscope to determine the extent of binding. On the other hand, the label can be an enzyme conjugated to the monoclonal antibody of the invention and the extent of binding can be determined by examining the slide for the presence of enzyme activity, which may be indicated by the formation of a precipitate, a color, or the like.

Another specific embodiment of the present invention involves the use of a surface to which the monoclonal antibody of the invention is attached. The underlying structure of the surface may take different forms, have different compositions and may be a mixture of compositions or laminates or combinations thereof. The surface may assume a variety of shapes and forms and may have varied dimensions, depending on the manner of use and measurement. Illustrative surfaces may be pads, beads, discs, or strips which may be flat, concave or convex. Thickness is not critical, generally being from about 0.1 to 2 mm thick and of any convenient diameter or other dimensions. The surface typically will be supported on a rod, tube, capillary, fiber, strip, disc, plate, cuvette and the like. The surface will typically be porous and polyfunctional or capable of being polyfunctionalized so as to permit covalent binding of the monoclonal antibody of the invention as well as to permit bonding of other compounds which form a part of a means for producing a detectable signal.

A wide variety of organic and inorganic polymers, both natural and synthetic, and combinations thereof, may be employed as the material for the slide surface. Illustrative polymers include polyethylene, polypropylene, poly(4-methylbuten), polystyrene, polymethacrylate, poly(ethylene terephthalat), rayon, nylon, poly(vinyl butyrate), silicones, polyformaldehyde, cellulose, cellulose acetate, nitrocellulose, latex, etc. Other materials which may be employed include paper, glasses, ceramics, metals, metaloids, semiconductor materials, cermits, silicates or the like. Also included are substrates that form gels, gelatins, lipopolysaccharides, silicates, agarose and polyacrylamides or polymers which form several aqueous phases such as dextrans, polyalkylene glycols (alkylene of 2 to 3 carbon atoms) or surfactants such as phospholipids.

The binding of the monoclonal antibody of the invention to the surface may be accomplished by well-known techniques, commonly available in the literature. See, for example, "Immobilized Enzymes," Ichiro Chibata, Halstad Press, New York (1978) and Cuatrecasas, J. Bio. Chem., 245:3059 (1970).

In carrying out the assay in accordance with this embodiment of the invention the sample is mixed with an aqueous medium and the medium is contacted with the surface containing the monoclonal antibody of the invention. Members of a signal producing system and any ancillary materials may also be included in the aqueous medium, either concurrently or added subsequently so as to provide a detectable signal associated with the surface. The means for producing the detectable signal can involve the incorporation in the medium of a labeled analyte or it may involve the use of a second monoclonal antibody having a label conjugated thereto. Separation and washing steps will be carried out as needed. The signal detected is related to the presence of an AIDS related infection. It is within the scope of the present invention to include a calibration as well as the measurement surface on the same support.

For an expanded discussion of the various general techniques discussed above with regard to conducting an assay on a surface see U.S. Patent Nos. 4,299,916; 4,391,904; 4,533,629; and 4,540,659.

Another example of a technique in accordance with the method and novel antibodies of the present invention is a non-flow cytometric technique. The sample to be analyzed and a monoclonal antibody of the present invention conjugated to a label are combined and incubated under conditions that will provide for agglutination when an AIDS related virus antigen is present in the sample. Desirable incubation times are about 10 to 600 seconds, preferably about 10 to 200 seconds at mild temperatures usually about 10 to 37°C. After incubation the medium is examined to determine any change in fluorescence as a result of agglutination. To this end one may use a non-flow cytometric technique in which a small diameter beam of light produced by means of slits or preferably a laser is used to differentiate particles based on their relative size. This technique employs fluorescent pulse height analysis or correlation of fluorescence fluctuations: Briggs, et al., "Homogeneous Fluorescence Immunoassay," Science, 212:1266-1267 (1981) and Nicoli, et al., "Fluorescence Immunoassay Based on Long-time Correlations of Number Fluctuations," Proc. Natl. Acad. Sci. USA, 7 (8):4904-4908 (1980).

A preferred method for determining a change of fluorescence in a non-flow-cytometric technique involves the use of a fiber optic cytometer described in U.S. Patent No. 4,564,598. Method and apparatus are disclosed for determining the presence of particles in a dispersion in relation to the detection of the presence or amount of a material of interest. An optical fiber is used to define a relatively small volume from which fluorescent light can be received and counted. The volume is related to the volume in which there is likely to be only a predetermined fluctuation. By employing a variety of techniques, which allow for changes in fluorescence fluctuations in relation to the presence of an analyte in the sample, the presence of the analyte can be determined. The fluctuations are observed over a period of time in a static mode or by sampling a plurality of volumes in the sample. A determination is then made by comparing the observed results obtained with solutions having a known amount of analyte.

Another specific embodiment of the present invention involves the use of the monoclonal antibodies of the invention in a "whole cell ELISA" test for the detection of HTLV-III expression on lymphocytes. Briefly, in a spectrophotometric procedure, peripheral blood lymphocytes are isolated by, for example, density gradient centrifugation and suspended in an aqueous protein containing medium (for example, an aqueous medium containing fetal bovine serum, glutamine, and RPMI/640). A small sample, for instance, 50µl, of the cell suspension can be added to wells of a 96-well microtiter plate and incubated, for example, for about 1 minute to 5 hours. After washing the cell-coated wells, a sample of an antibody of the invention is added to each well and incubated for 1 minute to 3 hours. After washing, chromogenic substrate, for example, orthophenylenediamine and hydrogen peroxide, is added to each well and incubated for 1 minute to 1 hour. Results can be read spectrophotometrically. In a microscopic procedure, buffered lymphocytes in suspen-

sion are incubated with antibody for 1 minute to 3 hours at a temperature of 0°C to 37°C. The cells are washed and then mixed with enzyme-labeled antibody for the above antibody for 1 minute to 3 hours at 0°C to 37°C. After washing, enzyme substrate is added and the cells are examined for the presence of a signal.

5 Another embodiment in accordance with the present invention involves the use of a Dot Blot technique to detect the presence of an antigen associated with an AIDS related viral infection. In such an approach a small sample, e.g., 1 µl, of patient serum is placed on a nitrocellulose strip next to a sample of normal serum. The strip is soaked in an aqueous buffered medium such as, e.g., phosphate buffered saline with gelatin, containing a monoclonal antibody of the invention, for a period of about 1 to 60 minutes, followed by
10 addition of labeled antibody for the monoclonal antibody.

In another embodiment in accordance with the present invention a flow cytometry-based detection system for HTLV-III virus on lymphocytes is employed. Flow cytometry means analysis by fluorescence-activated cell sorter (FACS). The principle of operation of flow cytometry have been reviewed in detail. See for example Bonner et al., Rev. scient. Instrum. (1972) 43:404-409; Herzenberg et al., Proc. Natl. Acad. Sci. U.S.A. (1979) 76:1453-1455; Miller et al., J. Immunol. Methods, (1981) 47:13-24; Loken et al., ibid (1982) 50:R85-R112; and Kruth, Anal. Biochem. (1982) 125 :225-242. A single cell suspension of the sample labeled with a fluorescent antibody is passed in single file through a narrow laser beam. The emitted fluorescence is collected by an optical system at right angles to the illumination. The emitted fluorescence of each individual cell is measured and stored and determination of the presence of the antigen in question
20 is made from the stored data.

Another aspect of the present invention involves the detection of a AIDS-related viral infection by the short term viral culture of clinical samples. The sample is cultured with H9 or CEM cells followed by detection utilizing the principles of flow cytometry, slide tests, or the like. "H9" means a T-cell lymphoma cell line. "CEM" means a T-cell lymphoma cell line. By "short term" is meant 2 to 48 hours, preferably 10
25 to 48 hours as compared to the present method which requires 20 to 30 days.

It is also within the scope of the present invention to employ the present methods and novel monoclonal antibodies in conjunction with screening test for the detection of antibodies to the AIDS-related viral antigens in serum. For example, patient samples can be screened utilizing a known antibody detection procedure. Patients identified as positive for the presence of such antibodies can be identified accordingly.
30 Samples taken from such patients can then be analyzed in accordance with the present invention to determine whether there is an AIDS-related viral infection present in such patient.

The invention also includes diagnostic kits for carrying out the method disclosed above. In the kit the reagents can be provided in packaged combination, in the same or separate containers so that the ratio of reagents provides for substantial optimization of a signal response to variations in analyte concentrations. In
35 one embodiment, the diagnostic kit can comprise (a) a monoclonal antibody more specifically defined above or a combination of such antibodies and (b) a conjugate of a specific binding partner for the above monoclonal antibody and a label capable of producing a detectable signal. The reagents may also include ancillary agents such as buffering agents and protein stabilizing agents, e.g., polysaccharides and the like. The diagnostic kit may further include, where necessary, other members of the signal producing system of
40 which system the label is a member, agents for reducing background interference in a test, control reagents, apparatus for conducting a test, and the like.

In another embodiment, the diagnostic kit can comprise a conjugate of a monoclonal antibody of the invention and a label capable of producing a detectable signal. Ancillary agents as mentioned above may also be present.
45

EXAMPLES

The invention is demonstrated further by the following illustrative examples.
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EXAMPLE 1Production of Monoclonal Antibodies to HTLV-III Antigens

5 Three month old female BALB/c and RBF/DN mice were immunized with preparations of HTLV-III antigen. The antigen preparation was a partially purified viral lysate derived from growth of HTLV-III in the human lymphoma, H9 cell line, obtained from the National Cancer Institute (NCI). Individual mice were immunized with 5-20 μ g antigen on polystyrene latex particles and boosted at weekly intervals with 10 μ g soluble antigen administered intravenously (I.V.) and intraperitoneally (I.P.). Sera were collected from the mice weekly intervals and subjected to Western Blot analysis for detection of antibody against HTLV-III viral antigens. Western Blot analysis showed the progressive development of antibodies to HTLV-III antigen over the 5 week immunization period. Mice showing reactivity to HTLV-III viral antigens were selected for fusions after 3, 4, and 5 weeks of immunization. Spleenocytes were fused to either the SP-2 or HL-1 myeloma cell lines using published procedures (for HL-1, Taggart, et al., *Science*, (1938) 219:1228; for SP-2, Schulman, et al., *Nature* (1978) 276:269). Screening procedures were selected to detect antibody in the 1-10 μ g/ml range. The initial screen was an ELISA procedure to detect immunoglobulin production followed by an ELISA screen to HTLV-III antigens (Table 1) which illustrates that the monoclonal antibodies are specific for the HTLV-III antigen. Positive wells were expanded and tested by Western Blot analysis. Positive cultures by Western Blot analysis were cloned and characterized. Hybrid continuous cell lines 3D8, 3F7, 3G12, 2A11, 4G2, and 1E2 were produced in this manner.

Several of the monoclonal antibodies expressed by the above cell lines detect multiple bands. This could indicate expression of shared epitopes on different proteins, detection of precursor molecules of differing molecular sizes or detection of a common carbohydrate determinant on distinct proteins. Treatment of the Western Blot strips with sodium periodate prior to reaction with monoclonal antibodies did not significantly alter the pattern of staining. This observation reduces the likelihood that carbohydrate epitopes are detected by the monoclonal antibodies. None of the anti-HTLV-III monoclonal antibodies react with normal lymphocytes or mast lymphocyte cell lines. However, three of them (1E2, 3F7 and 3G12 react with H9 cells. This suggests that the HTLV-III antigen preparation contained some H9 antigens and monoclonal antibodies to H9 proteins were produced.

EXAMPLE 2Detection of HTLV-III Antigen by Flow Cytometry

35 In order to evaluate the use of some of the monoclonal antibodies of the invention in detection of HTLV-III antigen expression on lymphocytes, latex beads (2.7 μ) were coated with HTLV-III antigen lysates and subjected to detection of the antigen by flow cytometry. Several of the antibodies detected the HTLV-III antigen as demonstrated by a shift in fluorescence intensity when a goat anti-mouse Ig-fluorescent second antibody reagent (TAGO, Burlingame, California) was employed (Table 2). These experiments demonstrated that using flow cytometry HTLV-III antigen could be detected on large particles using the monoclonal antibodies of the invention.

EXAMPLE 3

45 In additional experiments, the HTLV-III antigen was captured on latex beads using one of the above monoclonal antibodies. For this approach 1E2 or 3D8 was adsorbed to the latex beads and the beads were washed. The washed beads were then exposed to HTLV-III viral lysate, washed a second time and then mixed with a detection antibody. For detection, 3G12 was used (IgG₁ isotype). After washing, a fluorescent goat antimouse IgG, gamma chain specific, second antibody was reacted and the preparation evaluated by flow cytometry. As can be seen in Table 3, this capture and detection system using these two monoclonal antibodies is an effective means of identifying HTLV-III antigen. The monoclonal antibodies were also employed in reciprocal fashion so that 3G12 was used to capture the antigen and 3D8 was employed to identify the captured determinants.

EXAMPLE 4Detection of HTLV-III Antigen in the Blood of AIDS Patients

5 The double antibody capture technique described in Example 3 was then used to capture HTLV-III antigen from blood samples from patients with AIDS and from control patients. In a comparison study with 20 confirmed HTLV-III antibody positive samples, virus was detected in 12 samples. The samples were positive by Western Blot analysis but confirmation by direct viral culture was not carried out.

10 In additional experiments HTLV-III antigen was detected directly in clinical specimens by blotting the specimen onto nitrocellulose and detecting the antigen by a modified ELISA technique. In such an approach, 1 μ l of patient serum is placed on a strip of nitrocellulose next to 1 μ l of normal serum. The strip is then soaked in PBS-gelatin for 10 minutes followed by alkaline phosphatase conjugated goat anti-mouse antibodies in PBS-gelatin for 1 hour at room temperature. After three washes, a substrate consisting of bromochloroindole phosphate (BCIP)/nitro blue tetrazolium (NBT) is applied to the strip. Viral antigen 15 positive serum dots produce a purple dot within 10 minutes. Normal serum dots do not show any color development. The data in Table 4 show the results of these studies. Clones 3D8 and 3G8 detected HTLV-III antigen in the majority of samples from AIDS patients and AIDS positive control sera obtained from the National Cancer Institute (NCI). Direct comparison with virus culture was not carried out.

EXAMPLE 5Detection of HTLV-III Using a Whole Cell Technique

25 Using a modified whole cell ELISA technique HTLV-III antigen expression was detected on lymphocytes from patients who were confirmed positive by Western Blot analysis. In this test the cells were detected microscopically. Detection can also be made spectrophotometrically. In a spectrophotometric procedure, peripheral blood lymphocytes were isolated by density gradient centrifugation and suspended in RPMI 1640 (Standard tissue culture medium) with 2mM L-glutamine and 20% fetal bovine serum at a concentration of 2 30 x 10⁶ cells per ml. Fifty microliters of this cell suspension were added to a flat bottom well in a 96-well microtiter plate and incubated for about 3 hours. After washing the cell coated wells, anti-HTLV-III monoclonal antibody 3D8 was added to the well and incubated 1 hour. After washing, a citrate buffered orthophenylenediamine (OPD)/hydrogen peroxide substrate was added to each well and incubated 30 minutes. Positive color reactions were read by spectrophotometer. In the microscopic procedure, lymphocytes 35 were suspended in phosphate buffered saline with 0.1% gelatin and incubated with monoclonal antibody 3D8 for 1 hour at 4°C. The cells were then washed twice and then mixed with alkaline phosphatase conjugated goat anti-mouse IgG/IgM for 1 hour at 4°C. After washing, the cells were mixed with the substrate BCIP. HTLV-III positive cells were stained light to dark blue when viewed microscopically. The results are presented in Table 5.

EXAMPLE 6Detection of HTLV-III Expression on Lymphocytes by Flow Cytometry

45 HTLV-III attacks T4 lymphocytes and proliferates in those cells. Detection of HTLV-III antigen expression on T4 cells by flow cytometry using clones 3D8 and 3G12 was carried out. In this study samples were provided by the Oregon AIDS Task Force in blind fashion. The evaluations conducted included a monoclonal antibody panel consisting of the cell lines T₁, T_H, T_S, B and NK; a Western Blot analysis; T_H/T_S ratio; and the % of cells positive in the P_{pan} and T₄ categories to the above three anti HTLV-III monoclonals. 50 The patient groups included overt AIDS, ARC, PGL, and Western Blot positive but symptom free individuals. Controls consisted of high risk groups and low risk sex and age matched individuals. Eighty-two patients were evaluated. The data appears in Table 6. Forty-six of the samples came from patients with positive Western Blot analyses for HTLV-III antibody.

55 Monoclonal antibodies 3D8 and 3G12 detected expression of HTLV-III antigen on both T_{pan} and T₄ lymphocytes. There was a general correlation between disease severity and the number of positive cells. The lowest values in the patient groups were individuals who were as yet asymptomatic. The range of responses varied from 5% to 55% positive as a percentage of the T₄ cells with 3D8 or 3G12. By

comparison the high risk controls ranged by 0-5%. One control individual had a value of 7.5%. Interestingly this patient had an inverted T_H/T_S ratio although at this point was by Western Blot analysis negative. 3G12 resulted in higher background levels but also higher percentages of HTLV-III antigen expressing cells (see Table 6).

5 These data indicate that HTLV-III antigen expression can be effectively detected on T cells from Western Blot positive patients even in some cases where the patients are as yet asymptomatic. This may also be a means of detecting expression of the virus before antibody formation in high risk groups. Additional control groups in these studies included patients with Herpes 1 and 2, CMV and hepatitis. Patients in these viral control groups did not express antigens detected by the above monoclonal
10 antibodies.

EXAMPLE 7

15 Detection of HTLV-III Antigen Expression on H9 Cells Using Flow Cytometry

The H9 cell line is commonly used to detect HTLV-III antigen by culture and subsequent evaluation for the production of reverse transcriptase in the cells. To document that the monoclonals 3D8, 1E2 and 3G12 were actually detecting HTLV-III antigen on T_4 cells from patients, the following control experiment using H9
20 cells was conducted. H9 cells were grown with two virus containing preparations, either with a cell-free HTLV-III suspension or with T_4 lymphocytes that were expressing HTLV-III antigen. After 48 hours of culture the H9 cells were evaluated with monoclonal antibodies of the invention and control antibodies. As can be seen in Table 7 clones 3D8, 1E2, and 3G12 detected HTLV-III antigen expression on H9 cells only when the H9 cells had been infected. Infection with either cell free HTLV-III virus or HTLV-III positive T_4 cells was
25 equally effective; the isotype-control monoclonal antibody had no effect.

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TABLE 1

DETECTION OF HTLV-III ANTIGEN IN COMMERCIAL
SCREENING KITS

		<u>COMMERCIAL SCREENING KIT</u>	
<u>MONOCLONAL</u> <u>ANTIBODY</u>	<u>DILUTION</u>	<u>LITTON INDUSTRIES</u>	<u>ENI</u>
3F7	10 ⁻²	2.0*	2.0
	10 ⁻³	1.102	1.430
	10 ⁻⁴	0.43	0.620
3D8	10 ⁻²	2.0	2.0
	10 ⁻³	1.712	1.430
	10 ⁻⁴	1.252	0.821
3G12	10 ⁻²	1.862	2.0
	10 ⁻³	0.721	0.928
	10 ⁻⁴	0.210	0.286
2A11	10 ⁻²	0.641	0.589
	10 ⁻³	0.112	0.126
	10 ⁻⁴	0.05	0.06
1E2	10 ⁻²	0.761	0.682
	10 ⁻³	0.112	0.153
	10 ⁻⁴	0.05	0.05

*ELISA ABSORBANCE (in adsorbance units)

TABLE 2

FLOW CYTOMETRIC MEASUREMENT OF HTLV-III COATED BEADS
WITH ANTI-HTLV-III MONOCLONAL ANTIBODIES

<u>MONOCLONAL ANTIBODY</u>	<u>PERCENT BEADS STAINED</u>
3G8	60
4G2	18
3D8	82
2A11	77
3G12	73
3F7	75
1E2	80
Anti- β -hCG ^a	0.8
RPMI 1640 ^b	0.8

- a) antibody for β -human chorionic gonadotropin
b) standard tissue culture medium

TABLE 3

HTLV-III ANTIGEN DETECTION IN SERUM BY MONOCLONAL IgM ANTIBODY
CAPTURE - MONOCLONAL IgG ANTIBODY DETECTION USING LATEX BEADS

CAPTURE ANTIBODY ON BEAD	DETECTOR ANTIBODY	TEST SPECIMEN	% STAINED BEADS
1E2	3G12	Normal human serum (N.H.S.)	0.3
1E2	3G12	HTLV-III viral lysate ^a	66.4
3D8	3G12	N.H.S.	0.6
3D8	3G12	HTLV lysate	43.4
3D8	3G12	AIDS serum ^b	4.9

a) preparation of virus infected cells from cell culture, which cells are harvested, subjected to centrifugation, and lysed with detergent.

b) serum from patients having clinical AIDS.

TABLE 4

DETECTION OF HTLV-III BY MONOCLONAL ANTIBODY 3D8
USING A DOT BLOT TECHNIQUE

Total samples tested - 14

<u>SAMPLE CLASSIFICATION</u>	<u>NUMBER DOT BLOT POSITIVE</u>
Clinical AIDS	2/4
WESTERN BLOT POSITIVE	3/5
WESTERN BLOT NEGATIVE	0/5

TABLE 5

DETECTION OF HTLV-III BY A MICROSCOPIC WHOLE CELL
ELISA TECHNIQUE USING MONOCLONAL ANTIBODY 3D8

<u>SAMPLE CLASSIFICATION</u>	<u>PERCENT STAINED LYMPHOCYTES</u>
AIDS	19.0
AIDS	10.5
WESTERN BLOT POSITIVE	4.4
WESTERN BLOT POSITIVE	2.3
WESTERN BLOT POSITIVE	6.8
WESTERN BLOT POSITIVE	0.4
WESTERN BLOT POSITIVE	0.6

5

TABLE 6

10 DETECTION OF HTLV-III EXPRESSION ON T₄ LYMPHOCYTES
BY FLOW CYTOMETRY

15 GROUP	CONTROL	WESTERN BLOT POSITIVE	AIDS	ARC/PGL	ASYM ^c
Western Blot (+/-)	-	+	}	+	+
20 3D8/T4					
X ± SE	1.4 ± 0.3*	9.0 ± 4.2	}	25 ± 7	7.3 ± 1.4
RATIO:	$\frac{1}{35}$	$\frac{26}{20}$	}	$\frac{5}{1}$	$\frac{10}{5}$
25 #pos/#neg					
STATISTICS	p ^a <	0.0001		0.0001 0.05 p ^b <	0.0001 0.05 0.05
30 3G12/T4					
X ± SE	8.4 ± 2.8	28 ± 6.1	}	44 ± 10.1	29 ± 4.2
RATIO:	$\frac{1}{35}$	$\frac{32}{14}$	}	$\frac{5}{1}$	$\frac{12}{3}$
35 #pos/#neg					
STATISTICS	p ^b <	0.0001		0.0001 NS p ^b <	0.0001 0.06 NS

40

a Mann-Whitney U Test

b Fisher's Exact

45 c Asym - asymptomatic patients, i.e., those who were
sero-positive for AIDS antibodies but who had no overt
symptoms such as enlarged lymph nodes as found in ARC
or PGL patients or infectious diseases as found in
AIDS patients.

* number of positive cells detected by flow cytometry as
50 a percentage of 3D8 and 3G12 monoclonal antibodies on
T₄ lymphocytes.

55

TABLE 7

DETECTION OF HTLV-III EXPRESSION ON H9 CELLS

MONOCLONAL ANTIBODY	TREATMENT OF THE H9 CELL LINE		
	NONE	INFECTION WITH CELL FREE VIRUS	INFECTION WITH HTLV-III + T ₄ CELLS
SECOND ANTIBODY CONTROL	6.2*	6.4	3.1
3D8	7.5	18.5	29.7
1E2	14.9	40.8	41.9
3G12	32.2	82.8	94.0
ANTI-HCG CONTROL	3.4	5.6	4.6

*Percent cells positive by flow cytometry

Hybrid continuous cell lines 1E2, 3D8, and 3G12 were deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, MD 20852, under the Budapest Treaty on May 19, 1986, and received ATCC accession numbers HB9100, HB9101, and HB9102, respectively. Hybrid continuous cell lines 3G8 and 1C11 were deposited with the ATCC under the Budapest Treaty on May 28, 1986, and received ATCC accession numbers HB9114 and HB9115, respectively.

The invention has been described in detail with particular reference to the above embodiments. It will be understood, however, that variations and modifications can be effected within the spirit and scope of the invention.

Claims

1. A method for detecting the presence of HTLV-III infected cells in a medium which comprises contacting said medium with monoclonal antibodies against an antigen produced as a result of said infection and detecting the binding of said antibodies to said antigen with the proviso that, when said antigen is a viral gene product, said method involves detection of binding of said antibodies to said infected cells.
2. Method according to claim 1 wherein said medium is a human body fluid, preferably selected from whole blood, lymphatic fluid, serum, plasma, semen, saliva, and cerebral spinal fluid, or wherein said medium is a culture medium.
3. Method according to claim 1 or claim 2 wherein said antigen is a viral gene product or bound to a viral gene product.
4. Method according to claim 1 or claim 2 wherein said antigen is produced as a result of said infection and is not a viral gene product.
5. Method according to claim 4 wherein said antigen is bound to a lymphocyte, preferably wherein said lymphocyte is a T-helper cell, or to a macrophage.

6. Method according to claim 1 wherein said antigen is selected from the group of antigens characterised as molecules of approximately 24, 28, 39, 41, 55, 65, 49, and 52 kilodaltons, respectively.

7. Method according to any one of the preceding claims wherein said antibody is obtained from a hybrid continuous cell line selected from the group of hybrid continuous cell lines having the identifying characteristics of 3D8, 3F7, 3G12, 2A11, 4G2, 1E2, 3G8 and 1C11.

8. Method according to any one of the preceding claims wherein said antibody is conjugated to a label.

9. An immunoassay method for the detection of an AIDS virus related infection in a host, which comprises:

combining a body fluid from a host with a monoclonal antibody that binds to an antigen that is present only when said infection is present, and

examining the combination for the presence of immune complexes comprising said antigen and said monoclonal antibody with the proviso that, when said antigen is a viral gene product, said immune complex comprises said monoclonal antibodies and said antigen on infected cells.

10. Method according to claim 9 wherein said monoclonal antibody binds to a soluble antigen.

11. Method according to claim 9 or claim 10 wherein said AIDS virus related infection is selected from AIDS, ARC, PGL, and LAD.

12. An antibody capable of binding to an antigen produced as a result of a human HTLV-III infection, said antigen being a soluble serum antigen or an antigen bound to a lymphocyte, with the proviso that when said antigen is soluble, said antigen is not a viral gene product.

13. The antibody of claim 12 that is monoclonal, preferably wherein said monoclonal antibody is obtained from a hybrid continuous cell line having the identifying characteristics of 3D8, 1E2 or 3G12.

14. The monoclonal antibody of claim 13 conjugated to a label, which label is preferably selected from enzymes, radioisotopes, particles, supports, chromogens, chemiluminescers, fluorescers, coenzymes, free radicals and bacteriophages.

15. Monoclonal antibody according to claim 13 or claim 14 wherein said antigen is, or is bound to an HTLV-III viral gene product.

16. A diagnostic kit comprising in a packaged combination: a monoclonal antibody capable of binding to an antigen produced as a result of an infection by the viral causative agent of AIDS or a labelled derivative of said antibody, said antigen being bound to a viral gene product, or being bound to a lymphocyte as a result of said infection.

17. A composition of matter comprising a complex of:

(a) a monoclonal antibody that binds to a derivative of an HTLV-III gene product, and

(b) said gene product derivative.

18. A composition of matter comprising a complex of:

(a) a monoclonal antibody capable of binding to a cell-bound antigen other than an HTLV-III gene product and produced as a result of an HTLV-III infection, and

(b) said antigen.

19. A monoclonal antibody having the characteristics of a monoclonal antibody obtained from a hybrid continuous cell line having the identifying characteristics of 3D8, 3F7, 3G12, 2A11, 4G2, 1E2, 3G8 or 1C11.

Claims for the following contracting states: AT and ES

1. A method for detecting the presence of HTLV-III infected cells in a medium which comprises contacting said medium with monoclonal antibodies against an antigen produced as a result of said infection and detecting the binding of said antibodies to said antigen with the proviso that, when said antigen is a viral gene product, said method involves detection of binding of said antibodies to said infected cells.

2. Method according to claim 1 wherein said medium is a human body fluid, preferably selected from whole blood, lymphatic fluid, serum, plasma, semen, saliva, and cerebral spinal fluid, or wherein said medium is a culture medium.

3. Method according to claim 1 or claim 2 wherein said antigen is a viral gene product or bound to a viral gene product.

4. Method according to claim 1 or claim 2 wherein said antigen is produced as a result of said infection and is not a viral gene product.

5. Method according to claim 4 wherein said antigen is bound to a lymphocyte, preferably wherein said lymphocyte is a T-helper cell, or to a macrophage.

6. Method according to claim 1 wherein said antigen is selected from the group of antigens characterized as molecules of approximately 24, 28, 39, 41, 55, 65, 49, and 52 kilodaltons, respectively.

7. Method according to any one of the preceding claims wherein said antibody is obtained from a hybrid continuous cell line selected from the group of hybrid continuous cell lines having the identifying characteristics of 3D8, 3F7, 3G12, 2A11, 4G2, 1E2, 3G8 and 1C11.

8. Method according to any one of the preceding claims wherein said antibody is conjugated to a label.

9. An immunoassay method for the detection of an AIDS virus related infection in a host, which comprises:

combining a body fluid from a host with a monoclonal antibody that binds to an antigen that is present only when said infection is present, and

examining the combination for the presence of immune complexes comprising said antigen and said monoclonal antibody with the proviso that, when said antigen is a viral gene product, said immune complex comprises said monoclonal antibodies and said antigen on infected cells.

10. Method according to claim 9 wherein said monoclonal antibody binds to a soluble antigen.

11. Method according to claim 9 or claim 10 wherein said AIDS virus related infection is selected from AIDS, ARC, PGL, and LAD.

12. A method comprising production of an antibody capable of binding to an antigen produced as a result of a human HTLV-III infection, said antigen being a soluble serum antigen or an antigen bound to a lymphocyte, with the proviso that when said antigen is soluble, said antigen is not a viral gene product.

13. A method according to claim 12 wherein the antibody is monoclonal, preferably wherein said monoclonal antibody is obtained from a hybrid continuous cell line having the identifying characteristics of 3D8, 1E2 or 3G12.

14. A method according to claim 13 wherein the monoclonal antibody is conjugated to a label, which label is preferably selected from enzymes, radioisotopes, particles, supports, chromogens, chemilumescers, fluorescers, coenzymes, free radicals and bacteriophages.

15. Method according to claim 13 or claim 14 wherein said antigen is, or is bound to an HTLV-III viral gene product.

16. A diagnostic kit comprising in a packaged combinations:

a monoclonal antibody capable of binding to an antigen produced as a result of an infection by the viral causative agent of AIDS or a labelled derivative of said antibody, said antigen being bound to a viral gene product, or being bound to a lymphocyte as a result of said infection.

17. A composition of matter comprising a complex of:

(a) a monoclonal antibody that binds to a derivative of an HTLV-III gene product, and

(b) said gene product derivative.

18. A composition of matter comprising a complex of:

(a) a monoclonal antibody capable of binding to a cell-bound antigen other than an HTLV-III gene product and produced as a result of an HTLV-III infection, and

(b) said antigen.

19. A method comprising production of a monoclonal antibody having the characteristics of a monoclonal antibody obtained from a hybrid continuous cell line having the identifying characteristics of 3D8, 3F7, 3G12, 2A11, 4G2, 1E2, 3G8 or 1C11.

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(54) Method of detecting AIDS virus infection.

(57) A method is disclosed for detecting the presence of HTLV-III infected cells in a medium. The method comprises contacting the medium with monoclonal antibodies against an antigen produced as a result of the infection and detecting the binding of the antibodies to the antigen. The antigen may be a gene product of the HTLV-III virus or may be bound to such gene product. On the other hand the antigen may not be a viral gene product but may be produced as a result of the infection and may further be bound to a lymphocyte. The medium may be a human body fluid or a culture medium. A particular embodiment of the present method involves a method for determining the presence of an AIDS virus in a person. The method comprises combining a sample of a body fluid from the person with a monoclonal antibody that binds to an antigen produced as a result of the infection and detecting the binding of the monoclonal antibody to the antigen. The presence of the binding indicates the presence of an AIDS virus infection. Also disclosed are novel monoclonal antibodies, novel compositions of matter, and novel diagnostic kits.

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European Patent
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EUROPEAN SEARCH REPORT

Application Number

EP 87 30 3991

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
X	PROC. NATL. ACAD. SCI., vol. 82, no. 15, August 1985, pages 5199-5202, Washington, US; F. DI MARZO VERONESE et al.: "Monoclonal antibodies specific for p24, the major core protein of human T-cell leukemia virus type III" * Whole article *	1-3,6-19	G 01 N 33/569 G 01 N 33/577 C 12 P 21/00 // (C 12 P 21/00 C 12 R 1:91)
P,X	WO-A-8 604 423 (INSTITUTE OF CANCER RESEARCH) * Page 29, line 25 - page 30, line 7; claims 8,10,13 *	1-3,6-19	
P,X	EP-A-0 185 444 (CENTOCOR, INC.) * Page 5, lines 7-15; page 23, lines 8-20; claims 20-33 *	1-3,6-19	
P,X	WO-A-8 604 336 (INSTITUTE PASTEUR) * Page 2, lines 20-34; page 8, lines 2-10; claim 9 *	1-3,6-19	
A	WO-A-8 504 903 (UNITED STATES OF AMERICA) -----		TECHNICAL FIELDS SEARCHED (Int. Cl.4) C 12 P G 01 N A 61 K
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 07-02-1989	Examiner CARTAGENA Y ABELLA P.
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons ----- & : member of the same patent family, corresponding document			